

## The precious Ganoderma mushroom and plant diseases

Waill A. Elkhateeb\*, Ghoson M. Daba

Chemistry of Natural and Microbial Products Department, National Research Centre, Dokki, Giza, 12622, Egypt

**\*Corresponding Author:** Waill A. Elkhateeb, Chemistry of Natural and Microbial Products Department, National Research Centre, Dokki, Giza, 12622, Egypt.

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### Abstract

The oil palm, an economically important tree, has been one of the world's major sources of edible oil and a significant precursor of biodiesel fuel. Unfortunately, it now faces the threat of a devastating disease named basal stem rot disease. Many researchers have identified *Ganoderma boninense* and *Ganoderma zonatum* as the major pathogens that affects the oil palm tree and eventually kills. Identification of the pathogen causing basal stem rot disease is just the first step. No single method has yet been able to halt the continuing spread of the disease. This review focus on description and lifecycles of two of the major *Ganoderma* species pathogens on oil palm trees namely *G. boninense* and *G. zonatum*. Additionally, we highlighted some possible strategies to control these pathogens and limit the spread of basal stem rot disease.

**Keywords:** Mushrooms; *Ganoderma*; Plant diseases; *Ganoderma boninense*; *Ganoderma zonatum*.

### Introduction

Mushrooms have been used in many sides of human activity for many years. Some of these mushrooms have been called medicinal mushrooms due to their various morphological, physiological, and ecological characteristics that are also responsible for their diversity [1-25]. Some mushrooms and other fruiting bodies of filamentous fungi are edible and provide a good source of protein, whereas others have narcotic effects and used as medicine. Mushrooms contain many valuable secondary metabolites such as fatty acids, polysaccharides, phenolic compounds, and terpenoids [26-34]. Mushroom secondary metabolites especially from *Ganoderma* (White rot fungus belonging to Basidiomycetes) has been used for medicinal purposes for centuries particularly in China, Japan and Korea. A great deal of work has been carried out on *Ganoderma lucidum* have different biological activities including, anti-oxidant, anti-viral, anti-inflammatory, anti-coagulate, anti-cholesterol, anti-cancer, antimicrobial activities and other different activities. Mushroom-associated compounds, applied as therapeutic agents such as anti-aging and skin whitening agents. On the other hand, fungi in general and mushrooms in particular are famous producers of different enzymes of industrial applications. Such

enzymes have contributed in the efficient role of mushroom in bioremediation and recycling of elements in environment [35-40].

However, *Ganoderma* has its dark side which can cause major economic loss as it parasitizes on many economic important trees especially oil palm and coconut trees causing basal stem rot disease. Palm oil exports were valued at USD 23 billion in 2017 and USD 21 billion in 2018 [41]. The genus *Ganoderma* belongs to the family of Ganodermataceae within the Basidiomycetes, which causes white rots of hardwoods in many woody plants by decomposing lignin as well as cellulose and related polysaccharides. In the early stages of decay by *Ganoderma* spp., bleached zones usually appear in the wood, as a result of selective delignification. Elsewhere within the affected wood, there are less decolorised zones. These remain either relatively intact or show alterations typical of a simultaneous rot. The wood becomes progressively softer as the decay continues. Decay caused by *Ganoderma* spp. is often limited to the roots and the lower stem. The presence of *Ganoderma* spp., can usually be detected quite easily due to the development of the perennial fruit bodies. However, there are some species, for instance *Ganoderma resinaceum*, which

form only annual fruit bodies and which can therefore be easily overlooked. The fungal genus *Ganoderma* is a group of wood-decaying fungi that are found throughout the world on all types of wood; gymnosperms, woody dicots, and palms. [42]. In general, if *Ganoderma* a basidiocarp (conk) observe on a palm trunk, especially if it is still living, it is probably safe to assume it is *Ganoderma zonatum* and not some other *Ganoderma* species. Likewise, the *Ganoderma* species often observed on hardwood trees, such as oak, are rarely observed on living palms. These other *Ganoderma* species may occur on dead [43].

### ***Ganoderma* sp. infect oil palm at all stages**

*Ganoderma* sp. is an economically important plant pathogen especially in oil palm, causing basal stem rot disease. The distributions of *Ganoderma* species are worldwide in green ecosystem both to tropical and temperate regions [44]. The causal pathogen was originally identified as *Ganoderma lucidum*. In Malaysia, this causal pathogen was first recognised as *Ganoderma lucidum* [45]. The species *Ganoderma* has a wide host range, where more than 44 species from 34 genera of plants have been identified as potential hosts. Turner [46] reported fifteen species of *Ganoderma* from various parts of the world such as North America, Africa, India, Malaysia, Indonesia, Papua New Guinea and Thailand as being associated with BSR, including for example, *G. applanatum*, *G. boninense*, *G. chaliceum*, *G. cochlear*, *G. lucidum*, *G. miniatocinctum*, *G. pseudoferreum*, *G. tornatum*, *G. tropicum* and *G. zonatum*. Within these species *G. boninense*, *G. chaliceum*, *G. lucidum*, *G. miniatocinctum*, *G. pseudoferreum*, *G. tornatum* in isolates of diseased oil palms from different areas of Peninsular Malaysia, *G. boninense* is the most aggressive pathogen to cause the basal stem rot in oil palm [47].

The *Ganoderma* fungus is a facultative parasite that can live as a saprophyte on rotting stumps and roots, but when a suitable host (Oil palm). Subsequently, the disease spreads from plant to plant by roots or by spores [48, 49]. The bracket fungus, *Ganoderma boninense* Pat., causes basal stem rot (BSR) disease in oil palm plants (Figure, 1). Previously the disease was reported only in older age palms and currently found on young stages palms as well. Therefore, *Ganoderma* can infect all stages of oil palm plants. Disease symptoms only appear at the late stage of the disease and usually called a silent killer of oil palm. Progression of the disease is slow; however, it can destroy thousands of hectares of oil palm

plantations. Basal stem rot shortens the productive life of oil palms and causes serious economic losses to the oil palm industry. Hence, BSR is considered a serious threat to the oil palm industry in South East Asian countries. To date, there is high demand for sustainable detection and control of this disease. This review paper is elaborated on the detection and ecological impact of *Ganoderma* and causes basal stem rot disease in oil palms and plantation.

*Ganoderma* spp. can infect oil palm at all stages, from seedlings to old plants. Disease symptoms progress slowly, but usually every infected plant eventually dies. Usually, the disease develops from the roots but the external symptoms of BSR that appear on young palm shown as one-sided yellowing or molting of lower fronds, followed by necrosis [50]. New leaves become chlorotic, shorter, and the tips sometimes appear necrotic. Older plants when infected develop unopened whitish spear leaves. After a while, foliar symptoms become obvious as the parasitic fungus has already killed about half of the tissues of the plant. The roots of the infected palms are very friable, and their internal tissues become very dry and powdery. The cortical tissue is brown and disintegrates easily, and the stele becomes black in color [44]. Normally, infected young palms die within 6–24 months after these symptoms appeared, whereas mature palms take 2–3 years to die [44].

Elliott and Broschat, [43], reported that *Ganoderma* butt rot is caused by the fungus *Ganoderma zonatum*. This fungus attacks lignin in the position 4–5 feet lower of trunk and the trunk appears hard as the fungus does not cause a soft rot. All palms are considered hosts of *Ganoderma zonatum*. *Ganoderma zonatum* is not a primary pathogen of any other plant family. Wilting and decline was among reported symptoms and the disease is confirmed by detecting the basidiocarp (conk) on tree trunk before palm death. Basidiocarp is a shelf-like hard structure that appear attached to the lower palm trunk. Although, several diseased palms do not produce basidiocarp prior to death. A palm is diagnosed with *Ganoderma* butt rot by the formation of basidiocarp on the trunk, or internal discoloration of the trunk is detected after cutting the palm down. *Ganoderma zonatum* can be spread by spores that are produced then released from the conk. There are currently no cultural or chemical controls for preventing the disease or for curing the disease once the palm is infected [43].

### ***Ganoderma zonatum* description and life cycle**

*Ganoderma zonatum* is a white rot macrofungus that produces vast number of enzymes mainly lignin and



**Fig 1:** Different palm trees infected with different *Ganoderma zonatum*. Cited in: [www.srangetracking.com20180823butt-rot-in-palms](http://www.srangetracking.com20180823butt-rot-in-palms); [www.palmtalk.orgforumtopic70452-ganoderma-butt-rot-advice](http://www.palmtalk.orgforumtopic70452-ganoderma-butt-rot-advice)

cellulose that contributes in degrading woody tissue. *Ganoderma zonatum* is spread primarily by the spores produced in the basidiocarp (conk). The spores become incorporated into the soil, germinate and the hyphae then grow over the palm roots. The fungus does not rot the palm roots, it simply uses the roots as a means of moving to the woody trunk tissue. Once a palm is infected with *Ganoderma zonatum*, the fungus will move with that palm to the location in which it is transplanted [43]. As the fungus internally degrades the palm wood, the xylem will be affected at the end. Hence, the main symptom that may be detected is a sever to mild wilting of leaves except for the spear leaf. Also, basidiocarp is the remarkable structure associated with the fungus infection as the conk initiates from fungal growth inside the trunk. As the conk matures, a small shelf (bracket) will be formed as the basidiocarp starts to protrude from the trunk. A mature conk will appear in separate zones, hence came the name *Ganoderma zonatum*. The conk will appear in a half-moon shape with the relatively “straight” side directly attached to the trunk. *Ganoderma zonatum* basidiocarps, can reach 8 inches (at their widest point) and 2 inches thick. However, conks will take on the shape and size of the area in which they are growing. Basidiospores appear brownish-red and are produced in the pores on the underside of the conk. On the other hand, objects immediately around a conk that has dropped its spores may appear to be covered with a dust that is rusty in colour. One conk can produce 3 cups of spores [43].

#### *Ganoderma boninense* description and life cycle

*Ganoderma boninense* was recorded in Asia, Africa, and Central America. On oil palms, *G.*

*boninense* is recorded from Solomon Islands, Papua New Guinea, Australia and Samoa. Their hosts include oil palm, *Albizia*, *Casuarina*, betel nut, coconut, *Livistona*, and other palms. Nevertheless, it is difficult to specify hosts of *G. boninense* as different *Ganoderma* species appear similar. For example, in spite of attacking oil palms, *G. boninense* can be only identified by molecular methods, and the only method to test pathogenicity of samples is to put an infected rubber wood block under a seedling, and wait for numerous months (Figure, 2).

**Symptoms and Life Cycle;** In young palms, the lower leaves become yellow or develop yellow/green irregular spots and patches, called a mottle; they bend downwards, and then begin to rot prematurely. The unfolded leaves appear shorter than normal, yellowish in colour, and usually develop rots at the tips. Later, palms become pale and spear leaves remain unopened and growth slows. Similar symptoms were reported on mature palms as they also show numerous unopened spear leaves, and pale leaves. Moreover, dry rots happen internally in stem and bole, and powdery dry rots appear in roots. Within 6-12 months of symptoms appearance, young palms die, while mature palms can survive for about 2-3 years, or longer. The spore-producing structures or 'brackets', technically called 'basidiocarp' continue to form along the trunk, whether the palm remains in place, falls or is felled. They appear brown on top and white on bottom and are about 6 cm or greater. In some southern Asian countries, coconut is also infected by *G. boninense* causing leaves wilt, nuts become small, and lower leaves rot. Finally, a basal rot develops in the trunk, roots decay, and brackets develop [51].

The life-cycle of *Ganoderma boninense* is not well known. For instance, how do palms become infected? Is it from root-to-root contact, or from spores produced in the brackets? There is evidence for both. Support for root infections as the cause of basal stem rot comes from (i) artificial inoculations of seedlings, (ii) seedlings planted near *Ganoderma*-infected trunks in the field become infected the closer they are to the trunks, and (iii) the same is true for seedlings planted near *Ganoderma*-infected stumps from previous plantings. In these situations, roots become infected when they contact infected soil debris or infected roots. Spores are common in plantations, and so too are wounds made during routine harvesting - leaf bases are trimmed to release the fruit, and bunches are removed by cutting the fruit stalk - so it is possible spores infect oil palms at these times. Coconuts may also be a source of infection for

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oil palm after the coconuts have been cut down. There is also evidence that coconuts may be infected by *Ganoderma boninense* while alive but without showing symptoms until they die naturally and produce brackets [52, 53].



**Fig 2:** Different Palm trees infected with *Ganoderma boninense*. Cited in: [https://apps.lucidcentral.org/pppw\\_v10/text/web\\_full/entities/oil\\_palm\\_](https://apps.lucidcentral.org/pppw_v10/text/web_full/entities/oil_palm_)

### Control strategy of the disease

Basal stem rot disease caused by *Ganoderma* spp., considered most destructive disease in oil palm. This is because the disease escaped the early disease symptoms. Basal stem rot is like a 'cancer' in oil palm which is very difficult to detect at the early stages of the disease. In addition, resistant mycelium, basidiospores, chlamydospores, and pseudosclerotia at these resistance stages present in *Ganoderma boninense* and *Ganoderma zonatum* which are influence to inhibit control of *Ganoderma* [54].

Continuous research in combating the problem, the resolution remains stagnant. There are many control strategies for Basal stem rot include physical, chemical and biological procedures. Chemical treatment showed significant reduction in Basal stem rot occurrence when the oil palm trunk was injected with a combination of the fungicides carboxin and quintozone fungicides. However, chemical control agents are not good for the environment since they inhibit the growth of the other good microbes. Hence, growing concerns about the environment and the high cost of chemicals have encouraged farmers and researchers to find out the alternative way to control Basal stem rot, such as the use of biological control agents and pathogen-resistant cultivars [54].

In vitro studies have shown that the fungi *Trichoderma* spp., *Aspergillus* spp., and *Penicillium* spp. are antagonistic agents towards *Ganoderma*, (Figure, 3), [55]. When mass produced by antagonistic activity, these antagonists, especially

*Trichoderma* spp., are good bio-control agents for *Ganoderma* [56]. It is noted that *Trichoderma* can only protect the plants at the very early stages of the disease, and is not able to cure highly infected palms [57]. Even though, biocontrol agents have not provided a complete solution for Basal stem rot control, but the technique has become a notable approach compared to others. Hence, numerous bio-fungicides are currently available in the market for Basal stem rot control. Using biofungicide is a good agricultural practice for management of plant diseases. The most benefits of bio-fungicide reside in their biocompatibility and biodegradability that it controls the disease without causing any damage to the plant and leaving any toxic [58]. Using biofungicide is a good agricultural practice for management of plant diseases. The most benefits of bio-fungicide reside in their biocompatibility and biodegradability that it controls the disease without causing any damage to the plant and environment [59].



**Fig 3:** Green mold *Trichoderma* species on different *Ganoderma* fruiting body [53].

### Conclusion

Understanding the economic importance of oil palm and its contribution to world economy has rose a challenge to protect it against pathogens. There is no doubt that *Ganoderma* is an important medicinal mushroom, but also it causes the disease of oil palm and many other economically important trees. The two species causing majority of loss are *G. boninense* and *G. zonatum*. Studying the life cycle of these two species can contribute in finding effective strategies to face this problem with focus on biocontrol approaches. Further studies are required to improve potency of natural biocontrol agents such as the use of *Trichoderma* species. In order to protect economic importance trees.

### References

1. ALKolaibe, A. G., Elkhateeb, W. A., Elnahas, M. O., El-Manawaty, M., Deng, C. Y., Ting-Chi, W., & Daba, G. M. (2021). Wound healing, anti-pancreatic cancer, and  $\alpha$ -amylase inhibitory potentials of the edible mushroom,

- metacordyceps neogunnii. *Research Journal of Pharmacy and Technology*, 14(10), 5249-5253.
2. Elkhateeb, W., ELDien, A. N., Fadl, E., Elhagrasi, A., Fayad, W., & Wen, T. C. (2020). Therapeutic potentials of n-hexane extracts of the three medicinal mushrooms regarding their anti-colon cancer, antioxidant, and hypocholesterolemic capabilities. *Biodiversitas Journal of Biological Diversity*, 21(6).
  3. El-Hagrassi, A., Daba, G., Elkhateeb, W., Ahmed, E., El-Dein, A. N., Fayad, W., ... & Wen, T. C. (2020). In vitro bioactive potential and chemical analysis of the n-hexane extract of the medicinal mushroom, *Cordyceps militaris*. *Malaysian Journal of Microbiology*, 16(1).
  4. Elkhateeb, W., Elnahas, M., & Daba, G. (2021). Infrequent current and potential applications of mushrooms. In *Advances in Macrofungi* (pp. 70-81). CRC Press.
  5. Elkhateeb, W., Elnahas, M. O., Paul, W., & Daba, G. M. (2020). Fomes fomentarius and Polyporus squamosus models of marvel medicinal mushrooms. *Biomed. Res. Rev*, 3(1), 1-4.
  6. Elkhateeb W, Thomas P, Elnahas M, Daba G. (2021). Hypogeous and Epigeous Mushrooms in Human Health. *Advances in Macrofungi*, 7–19.
  7. Elkhateeb WA, Daba GM. (2022). Bioactive Potential of Some Fascinating Edible Mushrooms *Flammulina*, *Lyophyllum*, *Agaricus*, *Boletus*, *Letinula*, and *Pleurotus* as a Treasure of Multipurpose Therapeutic Natural Product. *Pharm Res*, 6(1): 1-10.
  8. Elkhateeb WA, Daba GM. (2022). Bioactive Potential of some Fascinating Edible Mushrooms *Macrolepiota*, *Russula*, *Amanita*, *Vovariella* and *Grifola* as a Treasure of Multipurpose Therapeutic Natural Product. *J Mycol. Mycological. Sci.*, 5(1): 1-8.
  9. Daba, G. (2020). The endless nutritional and pharmaceutical benefits of the Himalayan gold, *Cordyceps*; Current knowledge and prospective potentials. *Asian Journal of Natural Product Biochemistry*, 18(2).
  10. WA, E., Daba, G. M., & Soliman, G. M. (2021). The anti-nemic potential of mushroom against plant-parasitic nematodes. *J Microbiol Biotechnol*, 6(1).
  11. Elkhateeb WA, Daba GM, Elmahdy EM, Thomas PW, Wen TC, Mohamed N. (2019). Antiviral potential of mushrooms in the light of their biological active compounds. *ARC J Pharmac Sci.*, 5: 45–49.
  12. Elkhateeb, W. A., Daba, G. M., Elnahas, M. O., Thomas, P. W., & Emam, M. (2020). Metabolic profile and skin-related bioactivities of *Cerioporus squamosus* hydromethanolic extract. *Biodiversitas*, 21(10), 4732-4740.
  13. Elkhateeb, W. A., Daba, G. M., Elnahas, M. O., & Thomas, P. W. (2019). Anticoagulant capacities of some medicinal mushrooms. *ARC J Pharma Sci*, 5(4), 1-9.
  14. Elkhateeb, W. A., Daba, G. M., Thomas, P. W., & Wen, T. C. (2019). Medicinal mushrooms as a new source of natural therapeutic bioactive compounds. *Egyptian Pharmaceutical Journal*, 18(2), 88-101.
  15. Elkhateeb, W. A., & Daba, G. M. (2019). The amazing potential of fungi in human life. *ARC J. Pharma. Sci. AJPS*, 5(3), 12-16.
  16. Elkhateeb WA, Daba GM. (2020). Termitomyces Marvel Medicinal Mushroom Having a Unique Life Cycle. *Open Access Journal of Pharmaceutical Research*, 4(1): 1–4.
  17. Elkhateeb WA, Daba GM. (2021). Mycotherapy of the good and the tasty medicinal mushrooms *Lentinus*, *Pleurotus*, and *Tremella*. *Journal of Pharmaceutics and Pharmacology Research*, 4(3): 1–6.
  18. Elkhateeb, W. A., & Daba, G. M. (2021). The fascinating bird's nest mushroom, secondary metabolites and biological activities. *International Journal of Pharma Research and Health Sciences*, 9(1), 3265-3269.
  19. Elkhateeb WA, Daba GM. (2021). Highlights on the Wood Blue-Leg Mushroom *Clitocybe Nuda* and Blue-Milk Mushroom *Lactarius Indigo* Ecology and Biological Activities. *Open Access Journal of Pharmaceutical Research*, 5(3): 1–6.
  20. Elkhateeb WA, Daba GM. (2021). Highlights on the Golden Mushroom *Cantharellus cibarius* and unique Shaggy ink cap Mushroom *Coprinus comatus* and Smoky Bracket Mushroom *Bjerkandera adusta* Ecology and Biological Activities. *Open Access Journal of Mycology & Mycological Sciences*, 4(2): 1–8.
  21. Elkhateeb WA, Daba GM. (2021). Highlights on Unique Orange Pore Cap Mushroom *Favolaschia* Sp. and Beech Orange Mushroom *Cyttaria* sp. and Their Biological Activities. *Open Access Journal of Pharmaceutical Research*, 5(3): 1–6.
  22. Elkhateeb WA, Daba GM. (2022). Muskin the Amazing Potential of Mushroom in Human Life.

- Open Access Journal of Mycology & Mycological Sciences, 5(1): 1–5.
23. Elkhateeb WA, El Ghwas DE, Gundoju NR, Somasekhar T, Akram M, Daba GM. (2021). Chicken of the Woods *Laetiporus Sulphureus* and *Schizophyllum Commune* Treasure of Medicinal Mushrooms. *Open Access Journal of Microbiology & Biotechnology*, 6(3): 1–7.
  24. Elkhateeb WA, El-Ghwas DE, Daba GM. (2021). A Review on Ganoderic Acid, Cordycepin and Usnic Acid, an interesting Natural Compounds from Mushrooms and Lichens, 5(4): 1–9.
  25. Elkhateeb, W., Elnahas, M., Wenhua, L., Galappaththi, M. C. A., & Daba, G. M. (2021). The coral mushrooms *Ramaria* and *Clavaria*. *Studies in Fungi*, 6(1), 495-506.
  26. WA, E., Elnahas, M. O., Thomas, P. W., & Daba, G. M. (2020). *Trametes versicolor* and *Dictyophora indusiata* champions of medicinal mushrooms. *Pharm Res*, 4(1), 00019.
  27. Elkhateeb, W. A., Elnahas, M. O., Thomas, P. W., & Daba, G. M. (2019). To heal or not to heal? Medicinal mushrooms wound healing capacities. *ARC Journal of Pharmaceutical Sciences*, 5(4), 28-35.
  28. Elkhateeb WA, Karunarathna SC, Galappaththi MCA, Daba GM. (2022). Mushroom biodegradation and their role in mycoremediation. *Studies in Fungi*, (Under press).
  29. Elkhateeb, W. A. (2020). What medicinal mushroom can do. *Chem Res J*, 5(1), 106-118.
  30. Soliman, G., Elkhateeb, W., Wen, T. C., & Daba, G. (2022). Mushrooms as efficient biocontrol agents against the root-knot nematode, *meloidogyne incognita*. *Egyptian Pharmaceutical Journal*, 21(1), 68.
  31. Thomas, P. W., Elkhateeb, W. A., & Daba, G. (2019). Truffle and truffle-like fungi from continental Africa. *Acta mycologica*, 54(2).
  32. Thomas, P., Elkhateeb, W., & Daba, G. (2021). Industrial applications of truffles and truffle-like fungi. In *Advances in Macrofungi* (pp. 82-88). CRC Press.
  33. Daba, G. M., Elkhateeb, W. A., Wen, T. C., & Thomas, P. W. (2019). The continuous story of truffle-plant interaction. *Microbiome in Plant Health and Disease: Challenges and Opportunities*, 375-383.
  34. Zhang, X., Zhang, X., Gu, S., Pan, L., Sun, H., Gong, E., ... & Elkhateeb, W. A. (2021). Structure analysis and antioxidant activity of polysaccharide-iron (III) from *Cordyceps militaris* mycelia. *International Journal of Biological Macromolecules*, 178, 170-179.
  35. Elkhateeb, W. A., Daba, G. M., El-Dein, A. N., Sheir, D. H., Fayad, W., Shaheen, M. N., ... & Wen, T. C. (2020). Insights into the in-vitro hypocholesterolemic, antioxidant, antirotavirus, and anticolon cancer activities of the methanolic extracts of a Japanese lichen, *Candelariella vitellina*, and a Japanese mushroom, *Ganoderma applanatum*. *Egyptian Pharmaceutical Journal*, 19(1), 67.
  36. Elkhateeb, W. A., Daba, G. M., Sheir, D., El-Dein, A. N., Fayad, W., Elmahdy, E. M., ... & Wen, T. C. (2019). GC-MS analysis and in-vitro hypocholesterolemic, anti-rotavirus, anti-human colon carcinoma activities of the crude extract of a Japanese *Ganoderma* spp. *Egyptian Pharmaceutical Journal*, 18(2), 102-110.
  37. Elkhateeb, W. A., & Daba, G. M. (2022). Medicinal mushroom what should we know. *International Journal of Pharmaceutical Chemistry and Analysis*, 9(1), 1-19.
  38. Elkhateeb, W. A., EL-Ghwas, D. E., & Daba, G. M. (2022). Mushrooms as efficient enzymatic machinery. *J Biomed Res Environ Sci*, 3(4), 423-428.
  39. Elkhateeb WA, & Daba GM. (2022). The wild non edible mushrooms, what should we know so far?. *International Journal of Advanced Biochemistry Research 2022*; 6(1): 43-50.
  40. Zhang, X., Zhang, X., Gu, S., Pan, L., Sun, H., Gong, E., ... & Elkhateeb, W. A. (2021). Structure analysis and antioxidant activity of polysaccharide-iron (III) from *Cordyceps militaris* mycelia. *International Journal of Biological Macromolecules*, 178, 170-179.
  41. Purnomo, H., Okarda, B., Dermawan, A., Ilham, Q. P., Pacheco, P., Nurfatriani, F., & Suhendang, E. (2020). Reconciling oil palm economic development and environmental conservation in Indonesia: A value chain dynamic approach. *Forest Policy and Economics*, 111, 102089.
  42. Schwarze, FW, Ferner, D. (2003). *Ganoderma* on trees—differentiation of species and studies of invasiveness. *Arboricultural Journal*, 27(1): 59-77.
  43. Elliott, M. L., & Broschat, T. K. (2000). *Ganoderma* butt rot of palms. *University of*

*Florida IFAS Extension Bulletin*, 5.

44. Pilotti, C. A. (2005). Stem rots of oil palm caused by *Ganoderma boninense*: Pathogen biology and epidemiology. *Mycopathologia*, 159(1), 129-137.
45. Arrifin D, Idris AS, Singh G. (2000). Status of *Ganoderma* in oil palm. In Flood J, Bridge PD, Holderness M (eds) *Ganoderma diseases of perennial crops*. CABI Publishing: Wallingford.
46. Turner, P. D. (1981). *Oil palm diseases and disorders*. Oxford Univ. Press.
47. Wong, L., Bong, C. F. J., & Idris, A. S. (2012). *Ganoderma* species associated with basal stem rot disease of oil palm. *American Journal of Applied Sciences*, 9(6), 879-885.
48. Naher, L, Yusuf, UK, Ismail, A., Tan, SG, Mondal, MM. (2013). Ecological status of ' *Ganoderma*' and basal stem rot disease of oil palms ('*Elaeis guineensis*' Jacq.). *Australian Journal of Crop Science*, 7(11): 1723-1727.
49. Sanderson, F. R. (2005). An insight into spore dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia*, 159(1), 139-141.
50. Paterson, R. R. M. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop protection*, 26(9), 1369-1376.
51. Kandan, A., Bhaskaran, R., & Samiyappan, R. (2010). *Ganoderma*—a basal stem rot disease of coconut palm in south Asia and Asia pacific regions. *Archives of Phytopathology and Plant Protection*, 43(15), 1445-1449.
52. Rees, R. W., Flood, J., Hasan, Y., Wills, M. A., & Cooper, R. M. (2012). *Ganoderma boninense* basidiospores in oil palm plantations: evaluation of their possible role in stem rots of *Elaeis guineensis*. *Plant pathology*, 61(3), 567-578.
53. Chong, KP, Dayou, J, Alexander, A. (2017). Pathogenic nature of *Ganoderma boninense* and basal stem rot disease. In *Detection and control of Ganoderma boninense in oil palm crop* (pp. 5-12). Springer, Cham.
54. Hushiarian R, Yusof, NA, Dutse, SW. (2013). *Detection and control of Ganoderma boninense: strategies and perspectives*. Springer Plus, 2(1): 1-12.
55. Susanto, A., Sudharto, P. S., & Purba, R. Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159(1), 153-157.
56. Yan, Y., Zhang, C., Moodley, O., Zhang, L., & Xu, J. (2019). Green mold caused by *Trichoderma atroviride* on the lingzhi medicinal mushroom, *Ganoderma lingzhi* (Agaricomycetes). *International Journal of Medicinal Mushrooms*, 21(5).
57. Badalyan, S. M., GARibyan, N. G., & Innocenti, G. (2004). Interactions between xylotrophic mushrooms and mycoparasitic fungi in dual-culture experiments. *Interactions between Xylotrophic Mushrooms and Mycoparasitic Fungi in Dual-Culture Experiments*, 1000-1005.
58. Abdullah F, Ilias GN, Nelson M, Izzati NA, Yusuf KU. (2003). Disease assessment and the efficacy of *Trichoderma* as a biocontrol agent of basal stem rot of oil palms. *Science Putra*. 12: 31–33.
59. Chung, W. C., Huang, J. W., & Huang, H. C. (2005). Formulation of a soil biofungicide for control of damping-off of Chinese cabbage (*Brassica chinensis*) caused by *Rhizoctonia solani*. *Biological Control*, 32(2), 287-294.